

Research Article

Antifungal properties of ethanol extract of *vernonia amygdalina* stem and the effect on electrolyte status in normal and monosodium glutamate-intoxicated rats

Nicodemus E. Nwankwo^{1*}, Anthony C.C. Egbuonu², Priscillia O. Aliuba², and Emmanuel S. Okeke¹

¹Natural Science Unit, School of General Studies, University of Nigeria, Nsukka

²Department of Biochemistry, Michael Okpara University of Agriculture, Umudike, Abia, Nigeria

Corresponding Author:

N. E. Nwankwo

E-mail: : nicodemus.nwankwo@unn.edu.ng

Phone Number: +2348060428976

Received: 4 June 2019

Revised and Accepted: 26 June 2019

Abstract

This study investigated the antifungal properties of the extract of *Vernonia amygdalina* stem and its effect on the electrolyte status of normal and Monosodium Glutamate (MSG) intoxicated rats. In vitro antifungal property of the extract was tested. The serum electrolytes statuses of treated and non-treated MSG-intoxicated rats were evaluated. The results indicated the susceptibility of all the fungi used to the ethanol extract of *Vernonia amygdalina* stem. Extract inhibition of *Fusarium* spp showed a higher significant (* $p < 0.05$) zone of inhibition compared to ketoconazole. Serum chemistry showed that there was no significant difference (* $p < 0.05$) in the serum levels of bicarbonate, calcium and sodium at all doses tested. There was significant (* $p < 0.05$) increase in the serum potassium level at 400 mg/kg b.w. of the extract compared to the control. Significantly (* $p < 0.05$) raised sodium and calcium levels were observed in animals treated with 200mg/kg body weight. However, the elevated values were brought close to those of control at the dose of 400 mg/kg b.w. The studied extract exhibited considerable antifungal activity even against *Fusarium* spp. which the standard drug had no activity against. It was also found out that the administration of ethanol extract of *Vernonia amygdalina* stem did not alter the levels of serum electrolyte.

Keywords: Antifungal, Bitter Leaf, Electrolyte, Intoxication, MSG

1. INTRODUCTION

MSG has a diverse effect in animals [1] and adversely affected locomotor activities [2]. Adverse effect of MSG appears to manifest in MSG-sensitive individuals [3]. Lately, there is a shift from conventional and synthetic drug to natural products

like medicinal plants, in a quest to provide alternative drugs in alleviating various disease conditions. Medicinal plants are defined as those plants capable of producing active ingredients that treat several diseases and hence achieving an important role in global health [4]. The utilization of plants products and its waste has shown great potential for new drugs development and

correction of disease conditions [5]. Most of these plant wastes are disposed in such a way that they become source of environmental pollution [6].

Vernonia amygdalina commonly called ‘bitter leaf’ because of its bitter taste is a member of the *Asteraceae* family, ethno-medically consumed either as a vegetable or aqueous extract as tonics for the treatment of various illnesses. *Vernonia amygdalina* produces a variety of flavonoids and bitter sesquiterpene lactones which contribute to the bioactivities of this plant [7]. All parts of the plant are pharmacologically useful. Both the roots and leaves are used in phyto-medicine. In India, leaves, stem and root of *Vernonia amygdalina* are used in the treatment of HIV, measles, amoebiasis, influenza and mastitis infection [8].

Electrolytes are minerals in the body fluids that carry an electric charge. Electrolytes affect the amount of water, the acidity of blood (pH), muscle function and other important processes in the body [9]. Electrolytes are being regulated by the kidney, acid-base balance, insulin, aldosterone, gastrointestinal conditions and skin [10]. Administration of certain plant extracts such as *Picralima nitida* has been observed to tamper with the serum electrolytes levels [11]. The use of synthetic chemicals as fungicides cause severe and long-term environmental pollution, acutely toxic, and can even cause cancer in humans and other animals. Fungicides of biological origin have been demonstrated to be specifically effective on target organisms and are also biodegradable. Biological control has become popular worldwide. Thus this study was aimed at determining the antifungal properties of *Vernonia amygdalina* stem extract and its effect on the electrolyte status of normal and monosodium glutamate-intoxicated rats.

2. MATERIALS AND METHODS

2.1 Materials

Chemicals: All the chemicals used were of analytical grade and were product of Sigma Aldrich, St. Louis, USA.

2.2 Plant Material and Extraction: Fresh stems of *V. amygdalina* were collected from Amawom Oboro,

Ikwuano L.G.A in Abia State; the plant was identified and authenticated by Prof. H.O. Edeoga of the Department of Plant Science and Biotechnology, Michael Okpara University of Agriculture, Umudike. The dried stems were then taken for pulverization by a pulverizing machine in the Department of Soil Science, National Root Crops Research Institute, Umudike. A yield of 1.5 kg of the powder was obtained and 550 g of the powder was soaked in 2.2 litres of analyte ethanol for a period of 48 hours. The resulting mixture was then filtered using the Whatman’s filter paper. The filtrate obtained in a beaker of known weight was placed in a water bath at temperature of 40° C. After evaporation of the ethanol, the crude extract was refrigerated at 2- 8° C until use.

2.3 Experimental Animals: 20 adult male albino rats (*Ratus norregicus*) which weighed between 120–160 g were procured from the animal house of the Department of Biochemistry, College of Natural Sciences, Michael Okpara University of Agriculture, Umudike. They had free access to pelleted feed (Vital Feed) and clean water as they were allowed to acclimatize for two weeks.

2.4 Experimental Design: The rats were divided into five groups of four each as follows:

Group A: Rats administered MSG only

Groups B: Rats administered the extract only

Group C: Rats received only feed and clean tap water

Group D: Rats administered MSG and a low dose (200 mg/kg b.w.) of the extract

Group E: Rats administered MSG and a high dose (400 mg/kg b.w.) of the extract

Accurately weighed quantity of hydro-alcoholic (98 % ethanol) extract of *V. amygdalina* stem was suspended in distilled water and was administered orally to the experimental animals. MSG was freshly prepared in distilled water and administered orally with the aid of a gavage. Treatment was by daily intubation and lasted for 14 days.

2.5 Determination of Antifungal Activity:

Anti-fungal activity was evaluated using the agar well diffusion method as described by Magaldi *et al.* [12].

This was achieved by uniformly spreading 1 ml of fungi suspension prepared with sterile 0.85 % physiological saline solution on Saboureaud Chloramphenicol Actidione (SAC) plates. Afterwards, inoculum absorption SAC wells were made using sterile cork borers, which were then filled with 0.1 ml of the different concentrations *Vernonia amygdalina* stem extract was dissolved in distilled water (5 mg/ml). The control was carried out by filling the wells with 0.1 ml of mg/ml dissolved in sterile distilled water. The same procedure was followed for the determination of the diameters of inhibition of the antifungal agent ketonazole. Plates were incubated in steam room at 27°C. The results were read 5 days later. Standard diameter of inhibition was 25 ± 5. Every test was carried out in triplicate.

2.6 Determination of Serum Sodium Ion Concentration:

Serum sodium concentration was estimated using colorimetric method based on modified Maruna and Trinders method as described by Trinder [13].

To two test tubes labeled “test” and “standard” were added 1000 µl of precipitating reagent followed by the addition of sodium standard (10 µl) into the standard-labelled test tube and 10 µl of serum into the test-labelled test tube. The tubes were shaken vigorously and incubated for 5 minutes at room temperature and centrifuged at 2000 rpm for 2 min to obtain a clear supernatant. New test tubes; blank, standard and test were labelled and 1000 µl of sodium colour reagent was added to each of the tubes followed by the addition of standard supernatant (20 µl) into the standard-labelled tube and 20 µl of test supernatant obtained into the test-labelled tube; 20 µl of the precipitating reagent was added into the blank. The tubes were mixed and allowed to stand for 5 minutes at room temperature. The absorbance of standard and test against reagent blank was read and recorded

$$\begin{aligned} \text{Serum sodium concentration } \left(\frac{\text{mmol}}{\text{l}} \right) \\ = \frac{\text{Abs of B} - \text{Abs of T}}{\text{Abs of B} - \text{Abs of S}} \times \text{Conc. of standard} \end{aligned}$$

Where Abs = Absorbance

B = Blank, T = Test and S = standard.

2.7 Determination of Serum Potassium Ion Concentration: Serum potassium ion (K⁺) concentration was determined using the turbidometric method as described by Henry *et al.* [14].

Into two test tubes labelled standard and test were added 1000 µl of potassium reagent followed by the addition of standard (25 µl) into the standard-labelled tube and 25 µl of serum into the test-labelled tube. The tubes were shaken and incubated for 5 min at room temperature. Spectrophotometric reading was taken at 578 nm within 10 minutes. Serum potassium concentration was calculated as thus:

$$\begin{aligned} \text{Potassium concentration } \left(\frac{\text{mmol}}{\text{l}} \right) \\ = \frac{\text{Abs of test}}{\text{Abs of standard}} \times \text{Conc. of standard} \end{aligned}$$

2.8 Determination of Serum Calcium Ion Concentration: Serum calcium concentration was determined using the colorimetric method as described by Faulker and Meites [15].

Working reagent (1.0 ml) was added into test tubes labelled standard, control and test. Again, there was addition of 0.02 ml (10 µl) of samples to the respective tubes. The content was thoroughly mixed and allowed to stand for 60 sec at room temperature. The spectrophotometer was zeroed with blank at 570 nm and the absorbances of all tubes were read.

$$\begin{aligned} \text{Calcium concentration } \left(\frac{\text{mg}}{\text{dl}} \right) \\ = \frac{\text{Abs of test}}{\text{Abs of standard}} \times \text{Conc of standard} \end{aligned}$$

2.9 Determination of Serum Bicarbonate Ion Concentration: Serum bicarbonate ion was determined using enzyme spectrophotometric procedures as described by Forrester *et al.* [16].

To test tubes labelled blank, standard, control, and test were added 1.0 ml of CO₂ reagent and incubated for 3 minutes at 37° C. The spectrophotometer was zeroed with blank at 340 nm and maintained at 37° C. Water (0.005 ml) was pipette into the cuvettes labelled blank, standard and test respectively, followed by mixing via inversion and incubated for 5 minutes. The absorbances of all cuvettes were read at 340 nm.

$$\text{Bicarbonate content of sample} \left(\frac{\text{mmol}}{\text{l}} \right) = \frac{\text{Abs of B} - \text{Abs of test}}{\text{Abs of B} - \text{Abs of S}} \times \text{Conc of S}$$

2.10 Method of Statistical Analysis: The results were expressed as Mean \pm Standard Error of Mean (SEM) of replicate determinations and were subjected to one way analysis of variance (ANOVA), using Statistical Package for Social Sciences (SPSS-20) at 95 % level of confidence. Values were considered statistically significant at $p < 0.05$.

3. RESULTS

Data presented in table 1 below shows the antifungal

activity of the stem extract of *Vernonia amygdalina* when tested separately against the five test human pathogens. *A. niger* inhibition by varying doses of the extract were significantly ($*p < 0.05$) lower than antifungal activity of ketonazone. Inhibition of *H. capsulatum* by doses of the extract were significantly ($*p < 0.05$) lower than antifungal activity of ketonazone. Ketonazone showed higher significant ($*p < 0.05$) inhibition of *C. albicans* compared to graded doses of the stem extract. *P. notatum* inhibition by graded doses of the extract were significantly ($*p < 0.05$) lower than antifungal activity of ketonazone. Inhibition of *Fusarium spp* (4.0 ± 0.60) at 2000 mg/ml of extract *Vernonia amygdalina* stem showed a significant ($*p < 0.05$) increase compared to ketonazone.

Table1: Diameters of inhibition (mm) of *Vernonia amygdalina* stem extract on five fungal pathogens

| Conc. of stem extract | <i>A. niger</i> | <i>H. capsulatum</i> | <i>C. albicans</i> | <i>P. notatum</i> | <i>Fusarium spp</i> |
|-----------------------|-------------------|----------------------|--------------------|-------------------|---------------------|
| 2000 mg/ml | $10.0 \pm 0.58^*$ | $15.0 \pm 0.58^*$ | $21.7 \pm 0.88^*$ | $16.7 \pm 0.88^*$ | $4.0 \pm 0.58^*$ |
| 1000 mg/ml | $5.0 \pm 0.58^*$ | $7.7 \pm 0.33^*$ | $11.0 \pm 0.58^*$ | $7.7 \pm 0.88^*$ | $1.67 \pm 0.00^*$ |
| 500 mg/ml | $1.3 \pm 0.33^*$ | $3.0 \pm 0.58^*$ | $5.0 \pm 0.58^*$ | $3.0 \pm 0.58^*$ | 0.0 ± 0.00 |
| 250 mg/ml | $0.0 \pm 0.00^*$ | $1.0 \pm 0.58^*$ | $1.3 \pm 0.33^*$ | $0.6 \pm 0.33^*$ | 0.0 ± 0.00 |
| 125 mg/ml | $0.0 \pm 0.00^*$ | $0.0 \pm 0.00^*$ | $0.0 \pm 0.00^*$ | $0.0 \pm 0.00^*$ | 0.0 ± 0.00 |
| Ketonazone | 34.0 ± 0.58 | 26.7 ± 0.88 | 50.0 ± 1.15 | 28.0 ± 1.15 | 0.0 ± 0.00 |

Values are MEAN \pm Standard Error of Mean Triplicate determinations

Results in table 2 below show that there was no significant ($*p < 0.05$) difference in the bicarbonate levels between all the groups tested and the Normal control group. There was a significant ($*p < 0.05$) increase in potassium ion in all groups; MSG, extract, MSG+ Extract of *Vernonia amygdalina* stem (200 mg/kg and MSG+Extract of *Vernonia amygdalina* stem 400 mg/kg) (27.7 ± 0.50 , 28.1 ± 0.60 , 27.4 ± 0.50 and 28.4 ± 0.50) compared to normal control group (29.0 ± 0.80).The sodium level in the normal saline group (124.6 ± 2.70) was significantly ($*p < 0.05$)

higher than MSG, Extract, MSG+Extract of *Vernonia amygdalina* stem (185.5 ± 1.40 , 177.3 ± 1.20 , 185.3 ± 1.70 and 149.5 ± 0.70). There was a significant ($*p < 0.05$) increase in the calcium levels of the MSG group, Extract group and MSG+ Extract of *Vernonia amygdalina* stem 200 mg/kg (6.4 ± 0.50 , 5.5 ± 0.05 , 6.4 ± 0.80 and 4.6 ± 0.10) compared to group of normal saline (4.6 ± 0.10). The MSG+ Extract (400 mg/kg) was however non-significant ($*p > 0.05$) compared to the normal group.

Table 2: Effect of the extract on the electrolyte status of normal and monosodium glutamate-intoxicated rats

| GROUPS | HCO ₃ ⁻ | K ⁺ | Na ⁺ | Ca ²⁺ |
|-------------------------|-------------------------------|----------------|-----------------|------------------|
| MSG | 27.67 ± 0.54 | 5.62 ± 0.14 | 185.50 ± 1.44 | 6.43 ± 0.05 |
| EXTRACT | 28.09 ± 0.62 | 6.13 ± 0.05 | 177.25 ± 1.18 | 5.51 ± 0.05 |
| NORMAL | 29.02 ± 0.81 | 4.58 ± 0.03 | 124.60 ± 1.37 | 4.79 ± 0.22 |
| MSG+EXTRACT (200 mg/kg) | 27.42 ± 0.45 | 6.62 ± 0.05 | 185.25 ± 0.85 | 6.41 ± 0.08 |
| MSG+EXTRACT (400 mg/kg) | 28.44 ± 0.52 | 7.85 ± 0.80 | 149.50 ± 0.65 | 4.55 ± 0.13 |

Values are Mean ± S.E.M of quadruplicate determinations (n=4)

4. DISCUSSION

Results obtained during assay with the extract of *Vernonia amygdalina* stem showed its inhibitory effect at all concentrations against the five pathogenic fungi. All concentrations of the extract effectively suppressed the mycelial growth of these fungi and this effect was found to be dose-dependent. In a previous work by Fadina [17], ethanol and aqueous extracts of this plant at various concentrations inhibited the mycelial growth of the fungus *Colletotrichum lindemuthianum*. Two sesquiterpene lactones (*vernolide* and *vernodalol*) isolated from the leaves of *V. amygdalina* were found to inhibit the mycelial growth of some fungi [18].

Extract of *Vernonia amygdalina* stem caused decreased serum Na⁺. There was no significant difference in serum Na⁺ levels after administration of the ethanolic extract of *Vernonia amygdalina* stem and MSG. Even though the Na⁺ level increased significantly (*p < 0.05) at a low dose (200 mg/kg), it decrease at higher dose (400 mg/kg) compared to the control. This implies that the extract *V. amygdalina* stem possesses antihypertensive effect at a higher dose. The extract showed a significant effect on the calcium levels of the test animals compared to the control. It was observed that at 200 mg/kg b.w. of *V. amygdalina* extract, the calcium level was

significantly (*p < 0.05) reduced while at 400 mg/kg b.w., it was significantly (*p < 0.05) increased close to that of control; indication that a higher dose adversely alters the calcium electrolyte balance in the body. There was no significant (*p < 0.05) difference between the serum bicarbonate levels of the test groups compared to the control group. The ethanol stem extract of *Vernonia amygdalina* showed a significant effect on the potassium levels of the test animals as compared with control. It was observed that at 200 and 400 mg/kg b.w. of *V. amygdalina* extract, the potassium levels were significantly (*p < 0.05) compared to the control. This suggests a probable impairment of the kidney functionality due to administration of the extract.

5. CONCLUSION

The results of present study validated the traditional usage of the stem extract of *Vernonia amygdalina* against fungal infection. The extract of *V. amygdalina* stem did not have significant adverse effect on the functions of the kidney and maybe other organs of the body though the lowest dose caused significant perturbations in few serum electrolytes. Thus, the stem extract is a potential antifungal agent to be exploited by the pharmaceutical industry in the development of new antifungal drug with little or no adverse effects.

ACKNOWLEDGEMENT

We wish to acknowledge the staff members of Departments of Biochemistry and Plant Science and Biotechnology, Michael Okpara University of Agriculture, Umudike for their assistance during the period of this study.

REFERENCES

- [1] Egbuonu ACC, Obidoa O, Ezeokkonkwo XA, Ejikeme PM, Ezeanyika, LUS. Some biochemical effects of sub-acute oral administration of L-arginine on monosodium glutamate fed Wistar albino rats 1: Body weight changes, serum cholesterol, creatine, and sodium ion concentrations. *Toxicol Environ Chem* 2010; 92(7): 1331-1337.
- [2] Eweka AO, Om'iniabohs FAE. The Effects of *Monosodium glutamate* on the open field locomotor activities in adult Wistar rats. *Int J Nut Wellness* 2008; 6(2).
- [3] Egbuonu ACC, Osakwe ON. Effects of high monosodium glutamate on some serum markers of lipid status in male Wistar rats. *J Med & Med Sci* 2011; 2(1): 653-656.
- [4] Lima XF, Fenades-Ferreira M, Preiva-Wilson C. Pherolic Compounds protect Hep G2 cell from Oxidative Damage Relevance of Glutathione levels. *L. Fe Sci* 2006; 79: 2056-2068.
- [5] Maryam F, Mohammed EB. Evaluation of physicochemical properties of Iranian mango seed bernel oil, 2nd International Conference on Nutrition and Food Sciences IPCBCE 2013: 53-59.
- [6] Maisulthisakul P, Pasuk S, Rithiruangeles P. Antioxidant properties of phenolic phytochemicals from various cultivars of Thai mango seed kernels, *Asian J Food and Agro-ind* 2008; 1: 87-96.
- [7] Favi F, Cantrell CL, Mebrahtu T, Kraemer ME. Leaf peltate glandular trichomes of *Vernonia galamensis ssp. Galamensis vernonia, Ethiofica gilbert*: Development, ultrastructure, and chemical composition. *Int J Plant Sci* 2008; 169: 605-614.
- [8] Swee KY, Wan YH, Boon KB, Woon SL, Huynh K, Abdul HN, Neo-jahan BA. *Vernonia amygdalina*, an ethnoveterinary and ethnomedical used green vegetable with multiple bio-activities. *J Med Plants Res* 2010; 4(25): 2787-2812.
- [9] Hills J. Management of IV Fluids and electrolytes centre for Rural health (Rm, BSN, MSN (cancer), SpecertCR(Onc) 2013.
- [10] Ilegbedion IG, Onyide FM, Digba KA. Evaluation of MSG on Electrolyte Balance and Histology of Gastroesophageal Mucosa. *Middle-East J Sci Res* 2013; 18(2): 163-167.
- [11] Nwankwo NE, Egbuonu ACC, Nduka FO, Nwodo OFC. Effect of seed extract of *Picralima nitida* on haematological parameters of malaria-infected albino mice and its interference with the serum electrolyte levels. *Ife J Sci* 2017; 19(2): 379-388.
- [12] Magaldi S, Meta-Essayag S, Hartung de Capriles C, Perez C, Colella MT, Olaizola C, Ontiveros, Y. (2004). Well diffusion for antifungal susceptibility testing. *Int J Infect Dis* 2004; 8: 39-45.
- [13] Trinder P. A rapid method for the determination of sodium in serum. *Analyst* 1951; 76: 596.
- [14] Henry RF, Cannon DC, Winkelman JW. *Clinical chemistry principals and techniques*, 2nd Ed. Harper, and Roe, Hagerstown, MD 1974.
- [15] Faulker WR, Meister S. *Selected Methods for the Small Clinical Chemistry*, Washington D.C. 1982; 125 p.
- [16] Forrester RL, Wataji LJ, Silverman DA, Pierre KJ. Enzymatic method for determination of CO₂ in serum. *Clin Chem* 1976; 22: 243-245.
- [17] Fadina OO. The antifungal and nematicidal potentials of *Vernonia amygdalina* (Dal). *Acta Horticult* 2010; 853: 357-362.
- [18] Erasto P, Grierson DS, Afolayan AJ. Bioactive sesquiterpene lactones from the leaves of *Vernonia amygdalina*. *J Ethnopharmacol* 2006; 106(1): 117-120.